IgE-dependent sensitization in patients with COPD

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INTRODUCTION

Although it is well established that both asthma and COPD are immunologically driven conditions, their pathogenesis is not fully understood [1, 2, 3]. IgE-mediated sensitivity to inhaled allergens is strongly associated with asthma [3, 4]; however, this is not true for all asthma cases. There are also a few studies that suggest sensitization to environmental allergens also in COPD patients [5, 6].

Asthma is often considered to be a Th2–driven inflammatory disease, but many cases of the disease extend beyond a simple Th2-mediated disorder [3]. Altered immune responses also determine many pathobiological processes in COPD. Moreover, pro-inflammatory cytokines promote chronic airway inflammation in both asthma and COPD [1, 7].

OBJECTIVE

The aim of the study was to determine whether the presence and profile of IgE-dependent sensitization and/or the serum levels of some cytokines can differ between asthma and COPD.

MATERIALS AND METHOD

There were 351 patients who dropped out of the study. Screening was performed by a family doctor, a geriatrician, an internist, an allergist, or a trained nurse in outpatient clinics. Diagnosis of asthma and COPD was based on the GINA and GOLD guidelines, respectively [2, 3]. All patients had lung function tested by spirometry either prior to the study or during screening. Diagnosis of asthma was based on a medical history of asthma, physical examination, and partial reversal of airflow obstruction with a bronchodilator that resulted in a 12% or greater increase in FEV1 after inhalation of 400 mcg of salbutamol. Patients with episodic asthma were excluded. Healthy volunteers were recruited from the same outpatient clinics. These groups were matched with regard to age and gender and were consistent with the demographic structure of the Polish population over the age of 60. Additionally, 134 patients were excluded from the study because they did not fulfill the inclusion criteria. All patients signed consent forms to participate in the study which was approved by the Bioethics Committee. All of the centres obtained permission to publish the data collected.

The following patients were included in the study:

Patients with COPD. 103 participants (32 women and 71 men), age range: 41–79, mean age: 47.9± 9.1 years.
Inclusion criteria:
– men or women > 40 years of age;
– a current clinical diagnosis of COPD according to the GOLD guideline with COPD symptoms lasting more than one year;
– current or previous smoker with a smoking equivalent to 10 or more pack years;
– post-bronchodilator FEV₁/forced vital capacity (FVC) < 0.7 (70%) and 30% ≤ FEV₁ ≤ 70% of predicted normal value;
– any treatment of COPD;
– a score of dyspnea ≥ 2 according to the Modified Medical Research Council (MMRC).

Exclusion criteria:
– diagnosis of asthma or overlapping asthma/COPD syndrome;
– neoplastic disease or other unstable chronic diseases;
– any exacerbation of COPD during the six months prior to inclusion in the study (antibiotic or systemic steroids and/or hospitalization);
– severe unstable COPD and/or oxygen therapy because of a COPD exacerbation.

B. Patients with asthma. 114 participants (42 women and 72 men), age range: 69–77, mean 45.1 ± 5.8 years.

Inclusion criteria:
– men or women > 40 years of age;
– diagnosis of asthma according to the GINA criteria with a documented history of the disease lasting at least one year;
– 60% ≤ FEV₁ ≤ 80% of predicted normal value and a positive reversibility test defined as an increase in FEV₁ ≥ 12% and ≥ 200 ml relative to baseline after inhalation of 1 mg terbutaline;
– any treatment of asthma.

Exclusion criteria:
– diagnosis of overlap asthma/COPD syndrome;
– neoplastic disease or other unstable diseases;
– any exacerbation of asthma during the six months prior to inclusion in the study (antibiotic and systemic steroids and/or hospitalization);
– severe, difficult to control asthma.

C. Control group. 121 participants (46 women and 75 men), age range: 38–81, mean 44.8 ± 7.2 years.

Inclusion criteria:
– men or women > 40 years of age.

Exclusion criteria:
– diagnosis of COPD or asthma or overlapping asthma/COPD syndrome;
– neoplastic disease or any other unstable disease.

Characteristics of the control group, asthma patients and COPD patients are shown in Table 1.

The following procedures were performed in all patients.

Skin prick test. A skin prick test with inhaled allergens was performed according to published guidelines [8], using standardized protein extracts of D. pteronyssinus, D. farinae, grass mix, birch, alder, hazel, mugwort, Alternaria tennis, Cladosporium herbarium, Aspergillus fumigatus, cat, dog and cockroach (Allergopharma diagnostic set, Allergopharma, Reinbeck, Germany). A positive result was defined as a wheal > 3 mm in diameter, 20 min. after allergen extract application.

Serum levels of allergen-specific IgE. (sIgE) against the above-mentioned allergens were measured by ImmunoCAP (Pharmacia AB, Sweden), according to the manufacturer’s instructions. sIgE levels greater than 0.35 kUA/l were considered to be positive.

Immune analyses. Lymphocyte profiles were measured by the whole-blood method using fresh 10 ml blood samples that were treated with EDTA. The following lymphocyte markers were measured using ImmunoGEN kits (BD Bioscience, US) and a FACS Canto II flow cytometer (Becton Dickinson, US); CD3, CD29, CD16, CD56, CD4, CD8, and HLA-DR. The following lymphocyte markers were measured using the human Th1/Th2 cytokine kit II (ImmunoGEN, US) and a flow cytometer, as previously described. Cytokine concentrations are presented in pg/ml. Assay sensitivity for each cytokine were as follows: for IL-2, >31.2 pg/ml; IL-4, >2.1 pg/ml; IL-5, >12.5 pg/ml; IL-10, TNF, and IFN-gamma concentrations in the blood were measured using the human Th1/Th2 cytokine kit II (ImmunoGEN, US) and a flow cytometer, as previously described. Cytokine concentrations are presented in pg/ml. Assay sensitivity for each cytokine were as follows: for IL-2, >31.2 pg/ml; IL-4, >2.1 pg/ml; IL-5, >5.1 pg/ml; IL-10, >11.6 pg/ml; IL-13, >13.6 pg/ml; TNF >6.5 pg/ml; and IFN-gamma, >5.2 pg/ml.

Statistical analyses. Non-parametric statistical analyses were performed using the chi-square test and the multivariate ANOVA test. The risk of allergy symptoms was presented as a Hazard ratio. Spearman’s rank correlation test was used to examine any possible trends. The median and interquartile range were calculated for each cytokine. Statistica programme, version 8.0 (StatSoft, Poland) was used for all statistical analyses. A P value < 0.05 was considered to be statistically significant, following Bonferroni’s correction.

RESULTS

IgE-dependent sensitization was observed in 34 (33.3%) patients with a diagnosis of COPD. This was higher than the incidence of sensitization in the healthy subjects group (14 patients, 11.5%) but lower than that observed in asthma patients (46 patients, 40%). The sensitization profile for asthma patients was similar to that observed for COPD patients (Tab. 2). 32 patients with COPD were diagnosed with allergic rhinitis (sporadic in 14 patients and chronic
The serum concentration of IL-2 was significantly higher in both COPD and asthma patients than in healthy subjects (Tab. 3). When the subgroup of patients with non-allergic asthma was compared with patients with COPD, they were found to have similar serum concentrations of all cytokines, except for IFN gamma, which was lower in patients with COPD (Tab. 4).

### DISCUSSION

Both asthma and COPD are immunologically-driven disorders that are influenced by complex epigenetic agents [1, 9, 10]. It is commonly accepted that asthma is predominantly a Th2 cell-mediated inflammatory disease [3, 10], while COPD is associated with Th1 activity [1, 2]. IgE-mediated sensitivity to common aeroallergens is strongly associated with asthma pathogenesis [3]; however, the role of IgE-mediated sensitivity in COPD is less clear [11, 12].

In the presented study, atopy was associated with asthma, as expected, although atopic characteristics were also noted in COPD patients. Also as expected, sensitization to mites was observed in most asthmatics [3]. Hypersensitivity to Alternaria was also frequently observed in this group of patients. Nevertheless, sensitization to these allergens was also detected in those suffering from COPD, and significantly more frequently than in patients with non-allergic asthma or in the control group. These observations are in concordance with common knowledge about the role of airborne allergens in the pathophysiology of asthma, but airborne allergens are rarely believed to be associated with COPD [13, 14]. The similar sensitization profiles in patients with asthma and COPD observed in this study support the concept that asthma and COPD share some immunological features. The findings of the current study are in partial agreement with the results of Simpson et al. who demonstrated that Th1- and Th2-mediated disorders are positively correlated [15]. Moreover, these observations suggest a common origin for both commonly prevalent bronchoconstriction diseases.

The authors wish to emphasize that exposure to a variety of environmental factors plays a role in the development of these two immune diseases. The nature of these environmental factors, including allergens, smoking, and air pollution, determines whether asthma or COPD develops. Holt et al. suggest that allergic asthma, non-allergic asthma, and COPD are offshoots of a common ‘at-risk’ pathway underpinned by genotypes related to aberrations in control of host defence and tissue repair mechanisms [16]. They proposed that initiation of this pathway is programmed by the environmental experience of the immune system during infancy and early childhood, in particular by respiratory tract infections or sensitization to inhaled allergens in ‘at-risk’ subjects. The similar patterns in Th1/Th2 cytokines detected in patients with asthma and COPD in the presented

### Table 2. Comparison of the skin test results in the studied groups

<table>
<thead>
<tr>
<th>Allergens</th>
<th>COPD N=103</th>
<th>Asthma N=114</th>
<th>Control group N=121</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive results</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>D. pteronyssinus</td>
<td>29</td>
<td>62</td>
<td>9</td>
<td>0.01</td>
</tr>
<tr>
<td>D. farinae</td>
<td>23</td>
<td>59</td>
<td>8</td>
<td>0.01</td>
</tr>
<tr>
<td>Grass</td>
<td>16</td>
<td>48</td>
<td>7</td>
<td>0.01</td>
</tr>
<tr>
<td>Birch</td>
<td>6</td>
<td>19</td>
<td>6</td>
<td>0.01</td>
</tr>
<tr>
<td>Alder</td>
<td>8</td>
<td>18</td>
<td>4</td>
<td>0.03</td>
</tr>
<tr>
<td>Hazel</td>
<td>7</td>
<td>23</td>
<td>3</td>
<td>0.004</td>
</tr>
<tr>
<td>Mugwort</td>
<td>14</td>
<td>35</td>
<td>7</td>
<td>0.04</td>
</tr>
<tr>
<td>Alternaria</td>
<td>13</td>
<td>25</td>
<td>4</td>
<td>0.01</td>
</tr>
<tr>
<td>Cladosporium</td>
<td>8</td>
<td>31</td>
<td>2</td>
<td>0.009</td>
</tr>
<tr>
<td>Aspergillus</td>
<td>4</td>
<td>9</td>
<td>5</td>
<td>NS</td>
</tr>
<tr>
<td>Cat</td>
<td>6</td>
<td>15</td>
<td>8</td>
<td>0.03</td>
</tr>
<tr>
<td>Dog</td>
<td>2</td>
<td>3</td>
<td>0</td>
<td>NS</td>
</tr>
<tr>
<td>Cockroach</td>
<td>11</td>
<td>14</td>
<td>9</td>
<td>NS</td>
</tr>
</tbody>
</table>

* Multivariate ANOVA test; NS – not statistically significant

### Table 3. Th1/Th2 profiles in the studied groups

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>COPD N=103</th>
<th>Asthma N=114</th>
<th>Control group N=121</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Median with interquartile range (pg/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL-2</td>
<td>11.9;10.6–12.11</td>
<td>13.19;4.33–7.12</td>
<td>3.94;1.21–4.10</td>
<td>0.0001</td>
</tr>
<tr>
<td>IL-4</td>
<td>5.12;4.45–5.99</td>
<td>6.225.22–7.12</td>
<td>1.89;1.19–2.71</td>
<td>0.007</td>
</tr>
<tr>
<td>IL-5</td>
<td>3.25;2.02–4.11</td>
<td>3.51;2.21–4.14</td>
<td>2.91;2.11–3.41</td>
<td>NS</td>
</tr>
<tr>
<td>IL-10</td>
<td>21.81;18.33–21.61</td>
<td>23.01;22.11–24.81</td>
<td>10.38;9.44–12.82</td>
<td>0.014</td>
</tr>
<tr>
<td>TNF</td>
<td>14.09;13.76–15.32</td>
<td>16.15;15.09–17.03</td>
<td>9.12;8.56–10.01</td>
<td>0.022</td>
</tr>
<tr>
<td>IFN-gamma</td>
<td>7.74;6.55–8.12</td>
<td>6.14;5.42–7.31</td>
<td>3.45;2.27–4.07</td>
<td>0.014</td>
</tr>
</tbody>
</table>

* Multivariate ANOVA test; NS – not statistically significant
study support this notion. It is worth noting that both the asthma and COPD patients included in this study suffered from a moderate and stable form of the disease, with no exacerbations during the 12 months prior to inclusion in the study. However, in comparison with COPD patients, the asthma patients had more frequent family histories of allergy and a longer duration of the disease [16].

Both Th2 cells and Th1 cells express a panel of cytokines. IL-4, IL-10, IL-13 and GM-CSF are released by Th2 cells. Th1 cells are recognized by the expression and release of IL-2, IFN-gamma, and TNF-alpha [17, 18]. We evaluated a number of cytokines in our study to determine whether the polarization of the Th1/Th2 response at the serum level could differentiate asthma and COPD based on these data. The serum concentrations of all the cytokines evaluated in both asthma and COPD were similar, except at the lower level of IFN-gamma in COPD. This is important because the IFN-gamma plays a critical role in the innate and adaptive immune responses to viral and bacterial infections. IFN-gamma is predominantly produced by natural killer cells and by Th1 cells upon antigen-specific immune stimulation. There are data indicating that low IFN-gamma levels in serum suppress anti-tumour immunity [19]. IL-2 is a potent pro-inflammatory cytokine primarily released by Th0 and Th1 cells upon activation. IL-2 is commonly accepted to play a crucial role in the induction and maintenance of inflammatory processes in the airways [20, 21].

IL-4 has a dominant role in driving the differentiation of CD4+Th precursors into Th2 cells. It determines IgE synthesis by B lymphocytes and increases the expression of the low-affinity receptor for IgE (FceRII/CD23) to enhance antigen presentation by B cells. Moreover, inhalation of recombinant IL-4 has been shown to induce airway hyper-responsiveness and sputum eosinophils, providing evidence for the important role that this cytokine plays in the pathophysiology of asthma [22].

The similar cytokine profiles in asthma and COPD provide indirect evidence for the involvement of both Th1 and Th2 cells in the pathophysiology of these two disorders. IgE-dependent sensitization to airborne allergens has also been observed with comparable frequencies in asthma and COPD patients. It is worth noting that this was only the case for allergic asthma. In the presented study, more predominant atopy characteristics were observed in COPD than non-allergic asthma.

The results obtained reflect only a part of the complex network of immunologically-driven inflammatory processes in the airways of patients with bronchoconstriction diseases.

The inability of this study to differentiate between asthma and COPD patients based on their serum cytokine levels, indicates that neither Th2 nor Th1 related cytokines can be used as biomarkers for these disorders. Over-activation of either Th1 or Th2 responses occurs in both the pathogenesis of asthma and COPD. IgE-mediated sensitization plays a role not only in the pathogenesis of allergic asthma but also in the pathophysiology of COPD.

CONCLUSIONS

Neither IgE-dependent sensitization profiles to inhaled allergens, nor the profiles of Th1 and Th2 cytokine measured in patient serum, could differentiate between asthma and COPD patients with stable disease. The data obtained suggest that the pathophysiology of bronchoconstriction in asthma and COPD share some common features.

REFERENCES

3. Global Initiative for Asthma GINA. http://www.gina.org (accessed 22.03.15)