

ANTINUCLEAR ANTIBODIES ARE NOT INCREASED IN THE EARLY PHASE OF *BORRELIA* INFECTION

Radosław Śpiewak^{1,2}, Nimfa Maria Stojek¹, Jolanta Chmielewska-Badora¹

¹Department of Occupational Biohazards, Institute of Agricultural Medicine, Lublin, Poland

²Department of Dermatology, VU University Medical Centre, Amsterdam, The Netherlands

Śpiewak R, Stojek NM, Chmielewska-Badora J: Antinuclear antibodies are not increased in the early phase of *Borrelia* infection. *Ann Agric Environ Med* 2004, **11**, 145–148.

Abstract: In the literature, there are case reports suggesting that *Borrelia burgdorferi* infection may induce autoimmune diseases dependent on antinuclear antibodies (ANA). The present study was undertaken in order to verify this possibility in a prospective manner. The study group comprised 78 consecutive patients (51 women and 27 men, median age 41.5 years) referred to our Department for the serologic diagnosis of *Borrelia* infection. The patients' sera were tested for *Borrelia*-specific IgM and IgG (Recombinant Antigen Enzyme Immunoassays, Biomedica). Antibodies against *Borrelia* were detected in 31 (39.7%) persons. 15 persons (19.2%) had positive IgM, another 15 (19.2%) - positive IgG, and 1 person (3.2%) - both IgM and IgG. Frequent positivity of IgM antibodies suggests that persons in the early phase of infection prevailed in the group. Tests for anti-dsDNA, anti-RNP, anti-Sm antibodies, and a screening test for systemic rheumatic diseases (ANA Rheuma Screen) were carried out using Vareliisa Enzyme Immunoassays (Pharmacia&Upjohn). The spectrum of autoimmune diseases covered by these tests included SLE, MCTD, Sjögren's syndrome, scleroderma, polymyositis, and dermatomyositis. ANA were detected in 15 persons (19.2%): anti-dsDNA in 7 (9.0%), anti-RNP in 1 (1.3%), anti-Sm in 2 (2.6%), and ANA Rheuma Screen was positive in 6 persons (7.7%). Statistical analysis of differences in the ANA frequency between *Borrelia*-positive and -negative groups was carried out using Fisher's exact chi-square test (both without and with gender and age matching). No significant differences were found between the groups. Based on the above results, we conclude that there is no increase in the frequency of antinuclear antibodies in the early phase of *Borrelia* infection.

Address for correspondence: Dr hab. med. Radosław Śpiewak, Instytut Medycyny Wsi, ul. Jaczewskiego 2, 20-090 Lublin, Poland.
E-mail: spiewak@galen.imw.lublin.pl

Key words: *Borrelia burgdorferi*, borreliosis, Lyme disease, autoantibodies, antinuclear antibodies, anti-dsDNA, anti-RNP, anti-Sm, autoimmune diseases, serological study, Poland.

In the literature, there are casual observations suggesting that *Borrelia burgdorferi* infection may induce autoimmune diseases dependent on antinuclear antibodies (ANA), i.e. antibodies directed against the body's own cell nuclei. As infection by *B. burgdorferi* is common in Poland, this kind of complication would be of great importance to

public health. Until now, however, no scientific evidence has been produced for the existence and frequency of such complication. To fill this gap, the present study was undertaken in which we investigated the possible difference in the frequency of antinuclear antibodies between *Borrelia*-infected and non-infected people.

MATERIAL AND METHODS

Study population. A prospective study of consecutive patients referred to our department for the serologic diagnosis of *Borrelia* infection was carried out in autumn and winter 2001/2002. The criterion for inclusion was suspicion of borreliosis, clearly stated by the referring physician. Altogether, 78 persons were tested: 51 women and 27 men aged 9–78 (median 41.5) years.

Serology investigations. The patients' sera were tested for anti-*Borrelia* antibodies using IgM and IgG *Borrelia* Recombinant Antigen Enzyme Immunoassays (Biomedica Medizinprodukte, Vienna, Austria). At the same time, anti-dsDNA, anti-RNP, and anti-Sm antinuclear antibodies were sought in the serum by the means of Varelisa Enzyme Immunoassays, (Pharmacia&Upjohn, Freiburg). Additionally, a screening test for systemic rheumatic diseases was carried out using Varelisa ANA Rheuma Screen which includes the following nuclear antigens: dsDNA, histones, U1-snRNP, Sm, RNP-Sm, SS-A/Ro, SS-B/La, Scl-70, centromere, Jo-1, and PM-Scl-100. According to the manufacturer's information, the spectrum of autoimmune diseases covered by the serology tests includes SLE, MCTD, Sjögren's syndrome, scleroderma, polymyositis, and dermatomyositis. For all tests carried out, the qualitative results were interpreted according to the product information. In further statistical analysis, equivocal results were considered negative.

Statistical analysis. Depending on the positivity of *Borrelia* antibodies, the study subjects were divided into two groups denominated as *Borrelia*(+) and *Borrelia*(-) respectively. The frequencies of antinuclear antibodies in both groups were compared using Fisher's exact chi² test. As a second step, the gender and age matching of the study persons was carried out in order to avoid a possible bias caused by these factors [1, 22]. Groups consisting of the matched subjects were denominated as *Borrelia*(+)_{match} and *Borrelia*(-)_{match}. T-test for independent samples, Kruskal-Wallis test and the median test were used to compare age structures in the matched groups; none of these tests was capable of showing significant age difference. After the matching, the frequencies of antinuclear antibodies were re-analysed in matched groups using Fisher's exact chi² test (SPSS, Statsoft, Tulsa, USA).

RESULTS

Among 78 persons included into our study, antibodies against *Borrelia* antigens were detected in 31 (39.7%). 15 persons (19.2%) had positive IgM, another 15 (19.2%) - positive IgG; one person (3.2%) was both IgM- and IgG-positive. ANA were detected in 15 of the 78 persons (19.2%): anti-dsDNA in 7 (9.0%), anti-RNP in 1 (1.3%), and anti-Sm in 2 (2.6%). Varelisa Rheuma Screen was positive in 6 persons (7.7%). The frequencies of antinuclear antibodies in both groups are shown in Table 1. The

Table 1. Antinuclear antibodies in *Borrelia*-positive and negative groups: results before gender and age matching.

	N	ANA positive	anti-dsDNA positive	anti-RNP positive	anti-Sm positive	Rheuma Screen positive
<i>Borrelia</i> (+)	31	5 (16%)	2 (6%)	0	1 (3%)	2 (6%)
<i>Borrelia</i> (-)	47	10 (21%)	5 (11%)	1 (2%)	1 (2%)	4 (9%)

Table 2. Antinuclear antibodies in *Borrelia*-positive and negative groups: results after gender and age matching.

	N	ANA positive	anti-dsDNA positive	anti-RNP positive	anti-Sm positive	Rheuma Screen positive
<i>Borrelia</i> (+) _{match}	22	5 (23%)	2 (9%)	0	1 (5%)	2 (9%)
<i>Borrelia</i> (-) _{match}	22	6 (27%)	4 (18%)	0	0	3 (14%)

gender and age matching produced 12 pairs of women (median age for negative group 46 years, for positive group 45 years) and 10 pairs of men (for both groups median age was 37 years). The results for the matched groups are presented in Table 2. No significant differences were found both before and after matching.

DISCUSSION

Infection by *Borrelia burgdorferi* - a spirochete bacterium causing borreliosis (Lyme disease) is relatively common in Poland. Serological evidence of present or past *Borrelia* infection was found in 11–15% farmers in eastern Poland [4, 8] and in as many as 61% forestry workers in north-western Poland [25]. The main vector of the infection is *Ixodes ricinus* tick. A correlation has been observed between the tick activity and incidence of infection in exposed people [6]. *Borrelia* spirochetes could be detected in 8.8–13.2% of the ticks [3, 7, 31, 32, 40]. Other arthropods are also likely to transmit the infection, for example *B. burgdorferi* could be detected in up to 3.2% mosquitoes [21]. The situation in neighbouring Slovakia and Czech Republic is comparable to that in Poland [9, 41]. Domestic and wild animals form a large reservoir of the infection. For example, seropositivity to *B. burgdorferi* was found in 40% of dogs from endemic areas in Poland [29, 30], 17% of sheep and 19% of goats in Slovakia [37], and up to 60% of wild rodents in northern Czech territory [38].

Facing the high frequency of *Borrelia* infections, their immunological complications pose a serious threat to the public health. *Borrelia* was demonstrated to trigger a range of autoimmune reactions, i.e. reactions in which body's immune system turns against its own structures. The well-documented examples are inflammatory diseases of the central nervous system (CNS) and Lyme arthritis. In the CNS, *Borrelia* infection triggers an

Table 3. Overall frequency of antinuclear antibodies in the present study compared to other eastern-Polish populations (95%-confidence intervals are given in brackets).

	N	anti-dsDNA (%)	anti-RNP (%)	anti-Sm (%)	Rheuma Screen (%)	ReCombi ANA Profile* (%)
Present study	78	9.0 (2.6–15.3)	1.3 (0.0–3.8)	2.6 (0.0–6.1)	7.7 (1.8–13.6)	NT
Blood donors [34, 35]	50	2.0 (0.0–5.9)	2 (0.0–5.9)	0	NT	12.0 (3.0–21.0)
Mixed population [36]	130	9.2 (4.3–14.2)	4.6 (1.0–8.2)	1.5 (0.0–3.7)	NT	NT
Rural inhabitants [34]	90	12.2 (5.5–19.0)	5.6 (0.8–10.3)	2.2 (0.0–5.3)	NT	30.0 (20.5–39.5)

NT - not tested; *ReCombi ANA Profile covers 8 of 11 antigens present in Rheuma Screen - the results of both tests may be considered as comparable, though not identical.

autoimmune reaction that leads to demyelination of the nervous tissue [23]. Recently, an increased frequency of *Borrelia* antibodies among patients with multiple sclerosis was reported [5]. Lyme arthritis is an autoimmune reaction against human leukocyte function-associated antigen-1 (hLFA-1) which is cross-reactive with outer surface protein A (OspA) of *B. burgdorferi* [14, 15]. In this way, antibodies produced in order to destroy the invading bacteria bind and initiate damage to the body's own structures [28].

In contrast to the above-mentioned diseases, it remains unclear whether *Borrelia* infection is capable of inducing antibodies directed against the body's own cell nuclei (ANA). There are a handful of case reports on the co-existence of *Borrelia* infection and ANA-related autoimmune diseases, such as dermatomyositis [2, 12, 17, 19], systemic lupus erythematosus (SLE) [11] and generalized morphea [24]. Moreover, histopathologic similarities between borreliosis and SLE were pointed out [10, 16], and a raised titre of ANA in the course of borreliosis was reported [13, 26]. On the other hand, false-positive reactions to *Borrelia* antigen were described among patients with SLE [20, 39].

In the present study, we were not able to confirm any relationship between the presence of *Borrelia* antibodies and ANA. Of 31 *Borrelia*-positive persons, IgM-class antibodies were detected in 16, which indicates an early stage of the infection. It cannot be excluded that autoimmune reactions might start only in later stages, bearing in mind that *Borrelia*-related immune disturbances may persist over 10 years after infection [33]. A comparative study of patients with progressed borreliosis would shed additional light on the topic.

As shown in Table 3, the overall ANA-positivity rates among persons participating in the present study (both *Borrelia*-positive and -negative) were higher than among eastern-Polish blood donors studied previously [34, 35], which may result from the fact that all the blood donors were males. The frequency of ANA in males is lower than in females, possibly due to the lower estrogen levels [1]. Present results were most comparable to data from another random eastern-Polish population [36]. At the same time, the observed rates were lower than those seen previously among rural inhabitants [34]. Increased frequencies

of ANA among rural residents are attributed to their long-term exposure to pesticides [18, 27, 34].

CONCLUSION

Based on the above results, we conclude that there is no increase in the frequency of antinuclear antibodies in the early phase of *Borrelia* infection.

REFERENCES

- Ahmed SA, Hissong BD, Verthelyi D, Donner K, Becker K, Karpuzoglu-Sahin E: Gender and risk of autoimmune diseases: possible role of estrogenic compounds. *Environ Health Perspect* 1999, **107**(Suppl 5), 681-686.
- Arniaud D, Mattei JP, Pham T, Guis S, Baiada-Demoux AL, Serratrice J, Roux H: Coexistent dermatomyositis, relapsing polychondritis, and positive Lyme serology. A case-report. *Rev Rhum Engl Ed* 1997, **64**, 589-590.
- Bukowska K, Kosik-Bogacka D, Kuźna-Grygiel W: The occurrence of *Borrelia burgdorferi* sensu lato in the populations of *Ixodes ricinus* in forest areas of Szczecin during 2000-2001. *Ann Agric Environ Med* 2003, **10**, 5-8.
- Chmielewska-Badora J: Seroepidemiologic study on Lyme borreliosis in the Lublin region. *Ann Agric Environ Med* 1998, **5**, 183-186.
- Chmielewska-Badora J, Cisak E, Dutkiewicz J: Lyme borreliosis and multiple sclerosis: any connection? A seroepidemic study. *Ann Agric Environ Med* 2000, **7**, 141-143.
- Cisak E, Chmielewska-Badora J, Dutkiewicz J, Zwoliński J: Preliminary studies on the relationship between *Ixodes ricinus* activity and tick-borne infection among occupationally-exposed inhabitants of eastern Poland. *Ann Agric Environ Med* 2001, **8**, 293-295.
- Cisak E, Chmielewska-Badora J, Rajtar B, Zwoliński J, Jabłoński L, Dutkiewicz J: Study on the occurrence of *Borrelia burgdorferi* sensu lato and tick-borne encephalitis virus (TBEV) in ticks collected in Lublin region (eastern Poland). *Ann Agric Environ Med* 2002, **9**, 105-110.
- Cisak E, Chmielewska-Badora J, Zwoliński J, Dutkiewicz J, Patarska-Mach E: Ocena częstości zakażeń wirusem kleszczowego zapalenia mózgu i krętkami *Borrelia burgdorferi* wśród rolników indywidualnych na terenie Lubelszczyzny. *Med Pr* 2003, **54**, 139-144.
- Derdakova M, Halanova M, Stanko M, Stefancikova A, Cislakova L, Pet'ko B: Molecular evidence for *Anaplasma phagocytophilum* and *Borrelia burgdorferi* sensu lato in *Ixodes ricinus* ticks from eastern Slovakia. *Ann Agric Environ Med* 2003, **10**, 269-271.
- Duray PH, Steere AC: Clinical pathologic correlations of Lyme disease by stage. *Ann N Y Acad Sci* 1988, **539**, 65-79.
- Federlin K, Becker H: Borrelieninfektion und systemischer Lupus erythematoses. *Immun Infekt* 1989, **17**, 195-198.
- Fraser DD, Kong LI, Miller FW: Molecular detection of persistent *Borrelia burgdorferi* in a man with dermatomyositis. *Clin Exp Rheumatol* 1992, **10**, 387-390.

13. Goebel KM, Krause A, Neurath F: Acquired transient autoimmune reactions in Lyme arthritis: correlation between rheumatoid factor and disease activity. *Scand J Rheumatol Suppl* 1988, **75**, 314-317.
14. Gross DM, Forsthuber T, Tary-Lehmann M, Etling C, Ito K, Nagy ZA, Field JA, Steere AC, Huber BT: Identification of LFA-1 as a candidate autoantigen in treatment-resistant Lyme arthritis. *Science* 1998, **281**, 703-706.
15. Guerau de Arellano M, Huber BT: Development of autoimmunity in Lyme arthritis. *Curr Opin Rheumatol* 2002, **14**, 388-393.
16. Hejka AG, England DM, Schmitz JL, Schell RF: The pathologist's view of Lyme disease. *Wis Med J* 1989, **88**, 17-20.
17. Hoffmann JC, Stichtenoth DO, Zeidler H, Follmann M, Brandis A, Stanek G, Wollenhaupt J: Lyme disease in a 74-year-old forest owner with symptoms of dermatomyositis. *Arthritis Rheum* 1995, **38**, 1157-1160.
18. Holsapple MP: Autoimmunity by pesticides: a critical review of the state of the science. *Toxicol Lett* 2002, **127**, 101-109.
19. Horowitz HW, Sanghera K, Goldberg N, Pechman D, Kamer R, Duray P, Weinstein A: Dermatomyositis associated with Lyme disease: case report and review of Lyme myositis. *Clin Infect Dis* 1994, **18**, 166-171.
20. Keymeulen B, Somers G, Naessens A, Verbruggen LA: False positive ELISA serologic test for Lyme borreliosis in patients with connective tissue diseases. *Clin Rheumatol* 1993, **12**, 526-528.
21. Kosik-Bogacka D, Bukowska K, Kuźna-Grygiel W: Detection of *Borrelia burgdorferi* sensu lato in mosquitoes (Culicidae) in recreational areas of the city of Szczecin. *Ann Agric Environ Med* 2002, **9**, 55-57.
22. Manoussakis MN, Tzioufas AG, Silis MP, Pange PJ, Goudevenos J, Moutsopoulos HM: High prevalence of anti-cardiolipin and other autoantibodies in a healthy elderly population. *Clin Exp Immunol* 1987, **69**, 557-565.
23. Martin R, Ortlaf J, Sticht-Groh V, Mertens HG: Isolation and characterization of *Borrelia burgdorferi*-specific and autoreactive T-cell lines from the cerebrospinal fluid of patients with Lyme meningoradiculomyelitis. *Ann N Y Acad Sci* 1988, **540**, 449-451.
24. Nakashima T, Maeda M, Hayashi T, Kitamura K: A case of generalized morphea with a high titer of anti-*Borrelia burgdorferi* antibodies. *J Dermatol* 1999, **26**, 821-824.
25. Niścigorska J, Skotarczak B, Wodecka B: *Borrelia burgdorferi* infection among forestry workers - assessed with an immunoenzymatic method (ELISA), PCR and correlated with the clinical state of the patients. *Ann Agric Environ Med* 2003, **10**, 15-19.
26. Rose CD, Fawcett PT, Eppes SC, Klein JD, Gibney K, Doughty RA: Pediatric Lyme arthritis: clinical spectrum and outcome. *J Pediatr Orthop* 1994, **14**, 238-241.
27. Rosenberg AM, Semchuk KM, McDuffie HH, Ledingham DL, Cordeiro DM, Cessna AJ, Irvine DG, Senthilselvan A, Dosman JA: Prevalence of antinuclear antibodies in a rural population. *J Toxicol Environ Health A* 1999, **57**, 225-236.
28. Sigal LH: Immunologic mechanisms in Lyme neuroborreliosis: the potential role of autoimmunity and molecular mimicry. *Semin Neurol* 1997, **17**, 63-68.
29. Skotarczak B: Canine borreliosis - epidemiology and diagnostics. *Ann Agric Environ Med* 2002, **9**, 137-140.
30. Skotarczak B, Wodecka B: Molecular evidence of the presence of *Borrelia burgdorferi* sensu lato in blood samples taken from dogs in Poland. *Ann Agric Environ Med* 2003, **10**, 113-115.
31. Skotarczak B, Wodecka B, Cichocka A: Coexistence DNA of *Borrelia burgdorferi* sensu lato and *Babesia microti* in *Ixodes ricinus* ticks from north-western Poland. *Ann Agric Environ Med* 2002, **9**, 25-28.
32. Stańczak J, Racewicz M, Kubica-Biernat B, Kruminis-Łozowska W, Dąbrowski J, Adameczyk A, Markowska M: Prevalence of *Borrelia burgdorferi* sensu lato in *Ixodes ricinus* ticks (Acari, Ixodidae) in different Polish woodlands. *Ann Agric Environ Med* 1999, **6**, 127-132.
33. Stricker RB, Burrascano J, Winger E: Longterm decrease in the CD57 lymphocyte subset in a patient with chronic Lyme disease. *Ann Agric Environ Med* 2002, **9**, 111-113.
34. Śpiewak R, Stojek NM: Antinuclear antibodies among eastern-Polish rural inhabitants. *Ann Agric Environ Med* 2003, **10**, 207-209.
35. Śpiewak R, Stojek NM: Antinuclear autoantibodies in blood for transfusions - a possible risk source for recipients? *J Invest Dermatol* 2001, **117**, 795.
36. Śpiewak R, Stojek NM: Lupus erythematosus autoantibodies among patients without collagenosis symptoms. *J Invest Dermatol* 2001, **117**, 551.
37. Travnicek M, Stefancikova A, Nadzamova D, Stanko M, Cislakova L, Pet'ko B, Mardzinova S, Bhide MR: Seroprevalence of anti-*Borrelia burgdorferi* antibodies in sheep and goats from mountainous areas of Slovakia. *Ann Agric Environ Med* 2002, **9**, 153-155.
38. Vostal K, Zakovska A: Two-year study of examination of blood from wild rodents for the presence of antiborrelian antibodies. *Ann Agric Environ Med* 2003, **10**, 203-206.
39. Weiss NL, Sadock VA, Sigal LH, Phillips M, Merryman PF, Abramson SB: False positive seroreactivity to *Borrelia burgdorferi* in systemic lupus erythematosus: the value of immunoblot analysis. *Lupus* 1995, **4**, 131-137.
40. Wodecka B: Detection of *Borrelia burgdorferi* sensu lato DNA in *Ixodes ricinus* ticks in North-western Poland. *Ann Agric Environ Med* 2003, **10**, 171-178.
41. Zakovska A, Nejedla P, Holikova A, Dendis M: Positive findings of *Borrelia burgdorferi* in *Culex* (*Culex*) pipiens pipiens larvae in the surrounding of Brno city determined by the PCR method. *Ann Agric Environ Med* 2002, **9**, 257-259.