

**PREVALENCE OF ANTIBODIES TO *ENCEPHALITOOZON CUNICULI*
(MICROSPORIDIA) IN ANGORA GOATS – A POTENTIAL RISK OF INFECTION FOR
BREEDERS**

Lýdia Čisláková¹, Ivo Literák², Pavol Bálent, Vlasta Hipíková³, Mária Levkutová³,
Milan Trávníček³, Alexandra Novotná³

¹Department of Epidemiology, Faculty of Medicine, P. J. Šafárik University, Košice, Slovak Republic

²Faculty of Veterinary Medicine, University of Veterinary and Pharmaceutical Sciences, Brno, Czech Republic

³University of Veterinary Medicine, Košice, Slovak Republic

Čisláková L, Literák I, Bálent P, Hipíková V, Levkutová M, Trávníček M, Novotná A: Prevalence of antibodies to *Encephalitozoon cuniculi* (microsporidia) in Angora goats – a potential risk of infection for breeders. *Ann Agric Environ Med* 2001, **8**, 289–291.

Abstract: The presence of antibodies against *Encephalitozoon cuniculi* in Angora goats was detected by the method of indirect immunofluorescence (IFAT). The animals reacting at the titre 1: 64 and more were considered positive. Of the total number of 48 sera examined, 4 were positive at the titre 1: 32 and 2 were positive at the titre 1: 64. The occurrence of antibodies against *E. cuniculi* indicates that one of the causes of disorders in the reproductive cycle in Angora goats may be microsporidia *Encephalitozoon cuniculi*, and that these animals may be potential sources of infection for people.

Address for correspondence: Prof. Dr. Lýdia Čisláková, Faculty of Medicine, P. J. Šafárik University, Šrobárova 2, 04180 Košice, Slovak Republic.
E-mail: cislakova@pobox.sk

Key words: goats, Protozoa, Microsporidia, *Encephalitozoon cuniculi*, zoonosis, IFAT.

INTRODUCTION

Encephalitozoonosis is caused by an intracellular eukaryote protozoan parasite *Encephalitozoon cuniculi* [9], belonging to the phylum Microspora of the family Encephalitozoonidae [18]. The development of the parasite occurs only in living host cells with active metabolism (endothelial cells and macrophages) inside the parasitophorous vacuoles.

Encephalitozoon cuniculi infects a wide range of mammalian hosts, including rodents [24], rabbits [10], dogs [20, 21], blue foxes [14], cows [6], and nonhuman primates [25]. In goats, *Encephalitozoon cuniculi* (*Nosema cuniculi*) was first described by Khanna and Iyer [8]. *Encephalitozoon cuniculi* has been also described as an opportunistic pathogen in patients with decreased

resistance, e.g. in patients with AIDS [3, 23]. Recently, Mathis *et al.* [11] has found a rabbit *Encephalitozoon cuniculi* isolate, classified by Didier *et al.* [4] as strain I, in human urine, confirming the zoonotic character of the disease.

A characteristic feature of the infection induced by *Encephalitozoon cuniculi* in susceptible hosts is usually its chronic latent course, with significantly higher mortality in young and older individuals. Latently infected hosts, given immunosuppression factors, will develop an acute, clinically apparent disease [16].

The most common modes for transmitting infections with *Encephalitozoon* spp. are ingestion and inhalation. Transplacental transmission of infection may also play an important role in the epizootiology of encephalitozoonosis, especially in carnivores and rodents [13].

The Angora goats investigated for the presence of antibodies against *E. cuniculi* were from intensive goat breeding. The abortions, stillbirths and births of weak kids were observed in this breeding.

The reproductive disorders and abortions in goats may be caused by a number of factors – a metabolic derangement, e.g. iodine deficiency [1], wrong zoohygienic conditions, and also bacterial, viral and parasitic infectious agents. One of the infectious agents responsible for infertility and abortions of goats may also be microsporidia *Encephalitozoon cuniculi* from the kingdom Protista. It is also an important fact that goats infected by microsporidia *Encephalitozoon cuniculi* may pose a potential source of infection for people, especially for breeders attending these animals.

MATERIALS AND METHODS

Goats. Forty-eight dams of Angora goats, imported originally from Denmark, were examined. The goats were grazed in the summer periods of 1994 and 1995 and housed permanently in a barn with access to a walk in 1996 and 1997. The major health problem in 1994 and 1996 was a high incidence of reproductive failures, including abortions, stillbirths and births of weak kids.

Sera. Blood samples were withdrawn from the *vena jugularis*. The serum samples prepared were stored at -20°C until serological examination.

Antigen of *Encephalitozoon cuniculi*. *Encephalitozoon cuniculi* parasites were grown in E6 cells (VERO green monkey kidney cells) for the provision of spores. The infected cells were cultivated in modified RPMI 1640 medium, supplemented with 5% foetal calf serum and the addition of antibiotics and antimycotics (penicillin, streptomycin and amphotericin B). After lysis, the infected cells broke and spores were released into the medium from which they were separated by centrifugation at $400 \times g$ for 30 min. After rinsing in Percoll, they were again centrifuged and stored at 4°C.

The indirect immunofluorescence antibody test (IFAT). The IFAT method was used to determine specific anti-*Encephalitozoon cuniculi* antibodies. The method was performed according to Chalupský *et al.* [2].

Fresh suspension of *Encephalitozoon cuniculi* from tissue culture was placed on each well of a slide. The slides were air-dried for 24 h, then fixed in absolute acetone for 15 min and air-dried. The goats' sera tested were serially diluted beginning at 1:16 and ending at 1:256. Each of the wells on the slide was covered with 10 µl of rabbit anti-goat immunoglobulin plus fluorescein isothiocyanate conjugate (Sigma, Saint Louis, USA) of 1:160 dilution. After 30 min at 37°C the slides were washed and air-dried. They then were counter-stained with Evans blue and coverslips mounted with buffered glycerine.

The animals whose sera reacted at a dilution of 1:64 or higher were considered to be positive.

RESULTS

Of 48 serum samples from the Angora goats assayed by IFAT, 4 were positive at the titre 1:16 (8,3%) and 2 were positive at the titre 1:64, which represented 4.1%. In positive cases, spores of *Encephalitozoon cuniculi* were shown as oval, fluorescent formations of $1.5 \mu\text{m} \times 2.5 \mu\text{m}$. At the titre 1:128 and higher the animals were negative.

DISCUSSION

Encephalitozoon cuniculi, originally detected in rabbits [24] is an obligate intracellular protozoan parasite, invading various species of the animal kingdom, including the goat. It was the first mammal microsporidian cultivated *in vitro* [17].

E. cuniculi has been known as an agent causing encephalitozoonosis, a chronic disease, usually with asymptomatic progression [19] involving the central and peripheral nervous system, kidneys and ureters. In 1995, the zoonotic character of encephalitozoonosis was proved [3] and microsporidia became of interest not only for veterinary but also for human medicine.

In goats, *E. (Nosema) cuniculi* was found in the tubules of the kidney which showed changes associated with a focal chronic interstitial nephritis [8]. Similar findings were also observed in the kidneys of mice [5], rabbits [7], dogs [12] and cows [15] suffering from encephalitozoonosis. By IFAT, the *E. cuniculi* infection in goats was first described by Waller *et al.* [22] in Sweden.

In 1995, Reetz [15] published a report on the immunohistochemical evidence of the microsporidia antigens, probably *E. cuniculi*, in two cases of abortions in cattle in the seventh month of pregnancy. Microbiological examination of fetuses and foetal placentas did not reveal any of the usual bacterial or mycotic agents of abortion.

The results of our serological examination have confirmed the occurrence of antibodies against *E. cuniculi* in goats. In this breed there were observed abortions, stillbirths and births of weak kids. Although the percentage of positive reactions showing the presence of antibodies against antigens of *E. cuniculi* was not high (4,1%), it indicates that one of the possible causes of reproductive disorders may also be microsporidia *Encephalitozoon cuniculi*. Thus, breeders and veterinarians may be exposed during deliveries to the transmission of pathogens from infected animals. Breeders could also be infected by cleaning and tending the animals, because the transmission by contact by hand contaminated with urine containing spores of *E. cuniculi* was also described.

Acknowledgements

This study was supported by the Slovak Grant Committee, Grant No. 1/7022/20.

REFERENCES

1. Bíreš J, Bartko P, Weissová T, Matisiák T, Michna A, Bírešová M: Clinical and metabolic response of goats suffering from iodopenia to potassium iodine application. *Vet Med-Czech* 1996, **41**, 177–182 (in Slovak).
2. Chalupský J, Vávra J, Bedrník P: Detection of antibodies to *Encephalitozoon cuniculi* in rabbits by the indirect immunofluorescent antibody test. *Folia Parasitol* 1973, **20**, 281–284.
3. DeGrotte MA, Visvesvara G, Wilson ML, Pieniazek NJ, Slemenda SB, DaSilva AJ, Leitch GJ, Bryan RT, Reves R: Polymerase chain reaction and culture confirmation of disseminated *Encephalitozoon cuniculi* in a patient with AIDS: successful therapy with albendazole. *J Infect Dis* 1995, **171**, 1375–1378.
4. Didier ES, Vossbrinck CR, Baker MD, Rogers LB, Bertucci DC, Shadduck JA: Identification and characterisation of three *Encephalitozoon cuniculi* strains. *Parasitology* 1995, **111**, 411–421.
5. El Naas A, Revajová V, Letková V, Halánová M, Štefkovič M: Murine encephalitozoonosis and kidney lesions in some Slovak laboratory animal breeding centres. *Helminthologia* 1998, **35**, 107–110.
6. Halánová M, Letková V, Macák V, Štefkovič M, Halán M: The first finding of antibodies to *Encephalitozoon cuniculi* in cows in Slovakia. *Vet Parasitol* 1999, **82**, 167–171.
7. Horváth M, Horváthová A, Štefkovič M, Halánová M: Encephalitozoonosis of animals – pathologico-anatomical changes, diagnostics and therapy. *Slov Vet J* 1996, **21**, 63–67 (in Slovak).
8. Khanna RS, Iyer PKR: A case of *Nosema cuniculi* infection in a goat. *Indian J Med Res* 1971, **59**, 993–995.
9. Levaditi C, Nicolau S, Schoen R: L'agent etiologique de l'encephalite epizootique du lapin (*Encephalitozoon cuniculi*). *Comptes Rendus de l'Academie des Sciences, Paris* 1923, **89**, 984–986.
10. Levaditi C, Nicolau S, Schoen R: L'etologie de l'encephalite epizootique du lapin, dans ses rapports avec l'etude experimentale de l'encephalite lethargique *Encephalitozoon cuniculi*. (nov. spec.). *Ann Inst Pasteur, Paris* 1924, **38**, 651–711.
11. Mathis A, Michel M, Kuster H, Muller C, Weber R, Deplazes P: Two *Encephalitozoon cuniculi* strains of human origin are infectious to rabbits. *Parasitology* 1997, **114**, 29–35.
12. McInnes EF, Stewart CG: The pathology of subclinical infection of *Encephalitozoon cuniculi* in canine dams producing pups with overt encephalitozoonosis. *J S Afr Vet Assoc* 1991, **62**, 51–54.
13. Mohn SF, Nordstoga K, Dishington IW: Experimental encephalitozoonosis in the blue fox. Clinical, serological and pathological examination of vixens after oral and intrauterine inoculation. *Acta Vet Scand* 1982, **23**, 490–502.
14. Nordstoga K, Westbye K: Polyarteritis nodosa associated with nosematosis in blue foxes. *Acta Pathol Microbiol Scand* 1976, **84**, 291–296.
15. Reetz J: Microsporidien als Ursache von Aborten beim Rind. *Tierärztl Umschau* 1995, **50**, 550–554.
16. Schmidt EC, Shadduck JA: Murine encephalitozoonosis model for studying the host-parasite relationship of a chronic infection. *Inf Immun* 1983, **40**, 936–942.
17. Shadduck JA: *Nosema cuniculi*: *in vitro* isolation. *Science* 1969, **166**, 516–517.
18. Sprague V, Becnel JJ, Hazard EI: Taxonomy of phylum Microspora. *Crit Rev Microbiol* 1992, **18**, 285–395.
19. Stewart CG, Botha WS: The relationship in dogs between primary renal disease and antibodies to *Encephalitozoon cuniculi*. *J S Afr Assoc* 1978, **59**, 1921–1988.
20. Stewart CG, Botha WS, van Dellen AF: The prevalence of *Encephalitozoon* antibodies in dogs and an evaluation of the indirect fluorescent antibody test. *J S Afr Vet Assoc* 1979, **50**, 169–172.
21. Štefkovič M, Baranová D, Halánová M: Serological diagnosis of canine encephalitozoonosis in the Košice region. *Folia Vet* 1997, **41**, 113–115.
22. Waller T, Uggla A, Bergquist NR: Encephalitozoonosis and toxoplasmosis diagnosed simultaneously by a novel rapid test: the carbon immunoassay. In: *Proc. 3rd Int. Symp. Vet. Labor. Diagn.*, Ames, Iowa, USA 1983, 171–178.
23. Weber R, Bryan RT, Schwartz DA, Owen RL: Human microsporidial infections. *Clin Microbiol Rev* 1994, **7**, 426–461.
24. Wright JH, Craighead EM: Infectious motor paralysis in young rabbits. *J Exp Med* 1922, **36**, 135–140.
25. Zeman DH, Baskin GB: Encephalitozoonosis in squirell monkeys (*Saimiri sciureus*). *Vet Pathol* 1985, **22**, 24–31.