

The role of hepcidin and haemojuvelin in the pathogenesis of iron disorders in patients with severe malnutrition

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Przybyszewska J, Żekanowska E. The role of hepcidin and haemojuvelin in the pathogenesis of iron disorders in patients with severe malnutrition. *Ann Agric Environ Med*. 2014; 21(2): 336–338. doi: 10.5604/1232-1966.1108600

Abstract

Introduction and objective. The clinical consequences of malnutrition are multi-directional and result in dysfunctions of the majority of internal organs and systems. The results of recent studies suggest that a significant role is played by malnutrition in pathophysiology of iron homeostasis disorders, but the underlying mechanism is unclear. The study describes the potential role of hepcidin and hemojuvelin in the pathogenesis of disorders of iron metabolism during malnutrition.

State of knowledge. The participation of hepcidin in regulating iron homeostasis encompasses inhibiting the absorption of food iron from enterocytes and inhibiting the release of stored iron from the reticuloendothelial system cells. One of the factors that increases the post-translational level is the expression of hepcidin is IL-6. In studies focused on malnutrition it was observed that in persons with protein and energy deficits the level of proinflammatory cytokine, i.e. interleukin-6, in the serum, increased. The involvement of haemojuvelin in the overall iron homeostasis is related with the regulation of expression of the hepcidin coding gene on the transcription level. The highest haemojuvelin expression was observed in humans in skeletal muscles. Observations and analyses conducted *in vivo* allowed the conclusion that soluble HJV and cell-related haemojuvelin regulate hepcidin expression in response to changes in iron concentration. The research also demonstrated that soluble HJV neutralizes the inductive effect of IL-6 action on hepcidin expression.

Summary. It can be claimed that in persons with protein and/or protein-energetic malnutrition the muscle mass deficit may lead to insufficient production of haemojuvelin and sideropenia.

Key words

hepcidin, haemojuvelin, iron disorders, malnutrition

INTRODUCTION

Malnutrition is defined as pathology resulting from either prolonged deficit of energy nutrients and/or microelements in one's diet or recurring infections or chronic diseases [1]. Etiology differentiates three types of malnutrition:

- 1) energetic (marasmus, cachexia), developing due to exposure to long-term energy deficit;
- 2) protein (kwashiorkor), resulting from insufficient protein consumption and/or catabolic stress related with infections and extensive traumas;
- 3) mixed, i.e. protein energy malnutrition (PEM), most frequently observed in a variety of acute or chronic diseases [2].

While analyzing the causes of malnutrition, it should be borne in mind that especially in hospital patients it is of a secondary character and co-exists with many other diseases [3]. Reports also refer to cases in which the consequences of malnutrition are the initial symptom of another disease. Persons with high risk of developing severe malnutrition include patients suffering from cancer and gastrointestinal diseases, as well as psychological and neurological disorders [4, 5]. A high ratio of malnutrition occurrence is also reported

in patients with hyperthyroidism and in patients suffering from post-traumatic stress [4, 5].

The clinical consequences of malnutrition are multi-directional and result in dysfunctions of the majority of internal organs and systems [6]. The clinical picture of malnutrition shows skeletal muscle atrophy, proliferation of extracellular space with a tendency for the occurrence of oedema, immunity decrease, deficient wound healing, susceptibility to decubitus ulcers, decrease of physical activity, fatigue, apathy and hypothermia [4, 6, 7]. Results of research in the literature suggest that a significant role is played by malnutrition in the pathophysiology of iron homeostasis disorders.

Iron disorders in severe malnutrition. The unquestionable relation between malnutrition and iron disorders (manifested in microcytic anaemia) seems to be quite well reflected in a number of researches conducted among children with severe malnutrition [8, 9, 10]. The research show a significant increase of ferritin level in the serum of children suffering from malnutrition with concurrent iron deficit, in comparison to children with sideropenia who are not suffering from malnutrition. [8]. Ferritin concentration increase was additionally accompanied by a significant reduction of transferrin concentration and reduced TIBC (total iron-binding capacity). The research performed by Agarwali et al. [8] support the claim that malnutrition affects the development of iron disorders. Researchers showed that among such factors as infections, malnutrition, inflammation

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Received: 23 October 2012; accepted: 22 April 2013

or proteinuria, malnutrition exerted the biggest influence on the reduction of transferrin level and total iron-binding capacity [8]. Changes in protein plasma profiles probably result from the inflammatory process, co-occurring with malnutrition, in the course of which – at the expense of functionally important proteins, including transferrin – acute phase proteins, i.e., ferritin, are synthesized. The results of the aforementioned research seem to be particularly adverse in view of the research conducted by Broxmeyer et al. [11]. The authors research conducted *in vitro* showed the inhibitive influence of ferritin on the proliferation of myeloid cells [11]. Six years later, a team led by Vreugdenhil proved that removing excessive ferritin from the body by using iron chelating substances results in the improvement of haematological test results [12].

The fact that malnutrition may play a vital role in the pathophysiology of anaemia was also suggested by Mitrache et al. who relied on the results of research conducted on 186 patients with an average age of 85 (between 56–100 years of age) [13]. The authors showed in the multi-factor regression analysis that anaemia was significantly related with a low albumin level in the serum [13]. Other observations made in this study addressed the fact that 42% of geriatric patients with anaemia also met the criteria for malnutrition.

The role of hepcidin and haemojuvelin in the pathogenesis of iron disorders in severe malnutrition. Hepcidin is a peptide with high cysteine content and of molecular weight of ca. 3 kDa that occurs in body fluids in three forms. In the serum, isoforms of 25 and 20 amino acids are identified, and a peptide consisting of 25 amino acids is dominant [14]. Shorter forms i.e., made of 20 or 22 amino acids are identified mostly in urine [14, 15]. The mature and biologically active form of hepcidin contains in its sequence 8 cysteine residues connected by disulfide bonds [16]; disulfide bonds stabilise the cell structure [17]. Results of a number of researches focused on hepcidin show that it is a key mediator involved in iron homeostasis. The participation of hepcidin in regulating the overall iron homeostasis encompasses inhibiting the absorption of food iron from the lumen of absorptive enterocytes, and inhibiting the release of stored iron from the reticuloendothelial system cells. Nemeth et al., using cell cultures, showed that hepcidin performs its physiological activity through reactions with ferroportin (Fpn), i.e. the transmembrane protein receptor [18]. Ferroportin is present on the surface of cells, releasing iron to the circulating pool, such as absorptive enterocytes, macrophage, hepatocytes, and placenta cells [18]. The biological function of ferroportin is the transportation of iron from the cell interior to the blood vessels. Hepcidin is a factor regulating ferroportin expression on the post-translational level [18, 19]. It shows the capacity of binding directly with ferroportin molecules; hepcidin-Fpn binding is then subject to internalisation. Inside the emergent endosome, ferroportin is subject to lysosomal degradation. The loss of the cell membrane surface ferroportin is a secondary constraint on the process of iron release from the cell interior [18, 19].

Regulation of expression of the hepcidin coding gene (*HAMP*), located on the long arm of chromosome 19 (19q13), occurs both on the transcription and post-translational level [17, 20]. One of the factors with reported influence on hepcidin expression on the post-translational level is interleukin 6 [21–25]. Interleukin-6 induces hepcidin expression, thus

being a mediator in the development of hypoferrremia [23, 24]. In studies focused on malnutrition it was observed that in persons with protein and energy deficits (without any infection features) the level of proinflammatory cytokine, *inter alia* interleukin-6, in the serum increases [9, 10, 26]. A negative correlation between IL-6 and the condition of the muscle mass was also demonstrated [26]. The research results presented above also support the claim about the essential role of malnutrition in the pathogenesis of iron homeostasis disorders.

The involvement of haemojuvelin in the overall iron homeostasis is related with the regulation of expression of hepcidin coding gene on the transcription level. Activation of the hepcidin coding gene on the transcription level occurs through two varying cellular signalling pathways: through activating Stat3 and through activating Smad proteins [20, 27, 28]. Pro-inflammatory cytokines, in particular interleukin-6, induce the transcription process of the *HAMP* gene through activating Stat3, and then binding Stat3 to the regulatory region in the *HAMP* promoter [20, 27, 28]. The other mechanism of hepcidin expression control depend on the BMP/Smad signalling pathway. The connection of bone morphogenetic protein (BMP) with type II (BMPRII) or type I receptors (BMPRI) (BMP-typeI/typeII) entails phosphorylation of RSmad intracellular protein, which is then bound with Smad4 (also described as Co-Smad) [20]. The emerging complex (RSmad-Smad4) is relocated to the nucleus where, interacting with other transcription factors, it induces target genes, including the hepcidin gene [20].

The latest research shows a very important role played by haemojuvelin (HJV) in regulating the transcription of the *HAMP* gene through the BMP/Smad pathway [29–31]. Haemojuvelin is a protein product of the *HJV* gene located on the long arm of chromosome 1 (1q21). The highest haemojuvelin expression was observed in humans in skeletal muscles, the myocardium, liver, oesophagus and pancreas. Initially, researchers proved that in *in vitro* conditions haemojuvelin acts as a co-receptor of bone morphogenetic protein which facilitates the activation of the BMP-type I/type II complex [29]. Subsequent meticulous observations and analyses conducted *in vivo* allowed the conclusion that soluble HJV inhibits hepcidin expression [30, 31]. Intraperitoneal injection of recombinant soluble HJV into mice at the dose of 25mg/kg of bodyweight, three times per week for three consecutive weeks, resulted in the following: inhibition of hepcidin expression, increase in ferroportin expression and mobilization of stored iron from the reticuloendothelial system cells [30]. The final effect of the activity of exogenous soluble HJV was the iron increase in the serum [30].

The authors of the above-mentioned research also demonstrated that soluble HJV neutralizes the inductive effect of IL-6 action on hepcidin expression. Adding interleukin-6 into the hepatic cells (HepG2) culture increased hepcidin expression threefold [30]. The already observed and stimulating influence of IL-6 on the hepcidin synthesis was clearly neutralised when hepatocytes were incubated together with IL-6 and in combination with soluble HJV [30]. The research performed by Lin et al. demonstrated that soluble HJV and cell-related haemojuvelin jointly regulate hepcidin expression in response to changes in iron concentration in the extracellular space [31]. In view of the aforementioned biological activity of haemojuvelin, what seems particularly interesting are research results presenting increased release

of HJV by skeletal muscle cells in rats in response to iron deficits [32]. The authors of these reports suggest that skeletal muscles may perform a vital role in iron homeostasis [32]. It can be claimed that in persons with protein and/or protein-energetic malnutrition the muscle mass deficit may lead to insufficient production of haemojuvelin, thus yielding a secondary result of neutralising physiological response of the body to sideropenia.

Acknowledgement

This research received no specific grant from any funding agency in the public, commercial or non-profit sectors. J. P. was the major contributor. E. Ż. had a small but significant input.

There are no conflicts of interest.

REFERENCES

- WHO/NHD/007: Turning the tide of malnutrition: Responding to the challenge of the 21st century. World Health Organization, Geneva 2000.
- Management of severe malnutrition: a manual for physicians and other senior health workers. World Health Organization, Geneva 1999.
- DiMaria-Ghalili RA, Amella E. Nutrition in Older Adults: Intervention and assessment can help curb the growing threat of malnutrition. *AJN*. 2005; 105: 40–50.
- Sullivan DH, Bopp MM, Roberson PK. Protein-energy undernutrition and life-threatening complications among the hospitalized elderly. *J Gen Intern Med*. 2002; 17: 923–932.
- Thompson MP, Morris LK. Unexplained weight loss in the ambulatory elderly. *J Am Geriatr Soc*. 1991; 39: 497–500.
- Emery PW. Metabolic changes in malnutrition. *Eye*. 2005; 19: 1029–1034.
- Evans WJ. Protein nutrition, exercise and aging. *J Am Coll Nutr*. 2004; 23: 601–609.
- Agarwal MB, Mehta BC, Mhaiskar UM, Kumta NB, Shah MD. Effect of malnutrition on iron metabolism – a study of 45 children. *J Postgrad Med*. 1981; 27: 12–15.
- Dülger H, Arik M, Sekeroğlu MR, Tarakçıoğlu M, Noyan T, Cesur Y, Balahoroğlu R. Pro-inflammatory cytokines in Turkish children with protein-energy malnutrition. *Mediators Inflamm*. 2002; 11: 363–365.
- Sauerwein RW, Mulder JA, Mulder L, Lowe B, Peshu N, Demacker PN, Meer JW, Marsh K. Inflammatory mediators in children with protein-energy malnutrition. *Am J Clin Nutr*. 1997; 65: 1534–1539.
- Broxmeyer HE, Lu L, Bicknell DC, Williams DE, Cooper S, Levi S, Salfeld J, Arosio P. The influence of purified recombinant human heavy-subunit and light-subunit ferritins on colony formation in vitro by granulocyte- macrophage and erythroid progenitor cells. *Blood*. 1986; 68: 1257–1263.
- Vreugdenhil G, Nieuwenhuizen C, Swaak AJ. Interactions between erythropoietin and iron metabolism in anaemia of chronic disorders. *Eur J Haematol*. 1992; 48: 56–57.
- Mitrache C, Passweg JR, Libura J, Petrikos L, Seiler WO, Gratwohl A, Stähelin HB, Tichelli A. Anemia: an indicator for malnutrition in the elderly. *Ann Hematol*. 2001; 80: 295–298.
- Kemna EHJM, Tjalsma H, Podust VN, Swinkels DW. Mass spectrometry-based hepcidin measurements in serum and urine: analytical aspects and clinical implications. *Clin Chem*. 2007; 53: 620–628.
- Park CH, Valore EV, Waring AJ, Ganz T. Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem*. 2001; 276: 7806–7810.
- Hunter HN, Fulton DB, Ganz T, Vogel HJ. The solution structure of human hepcidin, a peptide hormone with antimicrobial activity that is involved in iron uptake and hereditary hemochromatosis. *J Biol Chem*. 2002; 277: 37597–37603.
- Krause A, Neitz S, Magert HJ, Schulz A, Forssmann W-G, Schulz-Knappe P, Adermann K. LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Lett*. 2000; 480: 147–150.
- Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DMcV, Ganz T, Kaplan J. Hepcidin Regulates Cellular Iron Efflux by Binding to Ferroportin and Inducing Its Internalization. *Science*. 2004; 306: 2090–2093.
- Knutson MD, Oukka M, Koss LM, Aydemir F, Wessling-Resnick M. Iron release from macrophages after erythrophagocytosis is up-regulated by ferroportin 1 overexpression and down-regulated by hepcidin. *Proc Natl Acad Sci U S A*. 2005; 102: 1324–1328.
- Domenico I, Ward DM, Kaplan J. Hepcidin regulation: ironing out the details. *J Clin Invest*. 2007; 117: 1755–1758.
- Kemna EHJM, Pickkers P, Nemeth E, van der Hoeven H, Swinkels DW. Time-course analysis of hepcidin, serum iron, and plasma cytokine levels in humans injected with LPS. *Blood*. 2005; 106: 1864–1866.
- Lee P, Peng H, Gelbart T, Beutler E. The IL-6- and lipopolysaccharide-induced transcription of hepcidin in HFE-, transferrin receptor 2-, and beta 2-microglobulin-deficient hepatocytes. *Proc Natl Acad Sci*. 2004; 101: 9263–9265.
- Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, Ganz T. IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest*. 2004; 113: 1271–1276.
- Nemeth E, Valore EV, Territo M, Schiller G, Lichtenstein A, Ganz T. Hepcidin, a putative mediator of anemia of inflammation, is a type II acute-phase protein. *Blood*. 2003; 101: 2461–2463.
- Nicolas G, Chauvet C, Viatte L, Danan JL, Bigard X, Devaux I, Beaumont C, Kahn A, Vaulont S. The gene encoding the iron regulatory peptide hepcidin is regulated by anemia, hypoxia, and inflammation. *J Clin Invest*. 2002; 110: 1037–1044.
- Visser M, Pahor M, Taaffe DR, Goodpaster BH, Simonsick EM, Newman AB, Nevitt M, Harris TB. Relationship of interleukin 6 and tumor necrosis factor-alpha with muscle mass and muscle strength in elderly men and women: the Health ABC Study. *J Gerontol A Biol Sci Med Sci*. 2002; 57: 326–332.
- Pietrangelo A, Dierssen U, Valli L, Garuti C, Rump A, Corradini E, Ernst M, Klein C, Trautwein C. STAT3 is required for IL-6-gp130-dependent activation of hepcidin in vivo. *Gastroenterology*. 2007; 132: 294–300.
- Verga Falzacappa MV, Spasic MV, Kessler R, Stolte J, Hentze MW, Muckenthaler MU. STAT-3 mediates hepatic hepcidin expression and its inflammatory stimulation. *Blood*. 2007; 109: 353–358.
- Babitt JL, Huang FW, Wrighting DM, Xia Y, Sidis Y, Samad TA, Campagna JA, Chung RT, Schneyer AL, Woolf CJ, Andrews NC, Lin HY. Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat Genet*. 2006; 38: 531–539.
- Babitt JL, Huang FW, Xia Y, Sidis Y, Andrews NC, Lin HY. Modulation of bone morphogenetic protein signaling in vivo regulates systemic iron balance. *J Clin Invest*. 2007; 117: 1933–1939.
- Lin L, Goldberg YP, Ganz T. Competitive regulation of hepcidin mRNA by soluble and cell-associated hemojuvelin. *Blood*. 2005; 106: 2884–2889.
- Zhang A-S, Anderson SA, Meyers KR, Hernandez C, Eisenstein RS, Enns CA. Evidence that inhibition of hemojuvelin shedding in response to iron is mediated through neogenin. *J Biol Chem*. 2007; 282: 12547–12556.